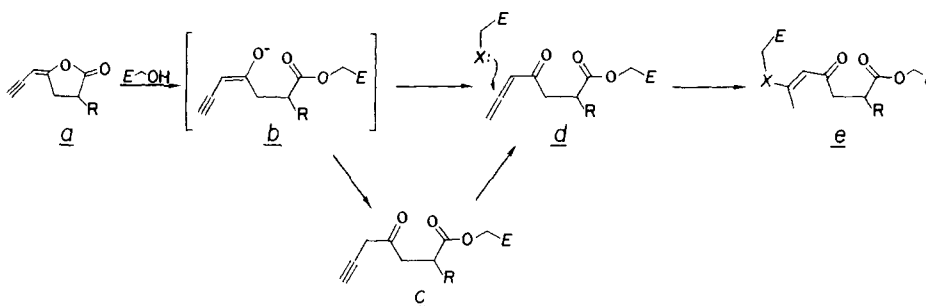


Scheme I



Scheme II

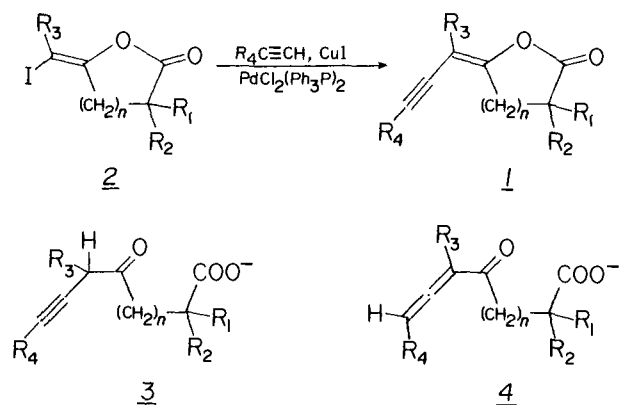


Table I

	R ₁	R ₂	R ₃	R ₄	n
a	H	H	H	H	1
b	<i>n</i> -Bu	H	H	H	1
c	PhCH ₂	H	H	H	1
d	PhCH ₂	H	H	Me	1
e	PhCH ₂	H	Me	H	1
f	PhCH ₂	Me	H	H	1
g	PhCH ₂	H	H	H	2

as suicide inhibitors. Three conditions, (1) acylation of the enzyme (a → b), (2) unmasking of the allene (b → d), and (3) capture of an enzyme nucleophilic residue, must be met in order to inactivate the enzyme.

(*E*)-Ynenol lactones **1** are conveniently synthesized⁵⁻⁷ by coupling the corresponding (*E*)-iodo enol lactones^{4c} **2** with the appropriate acetylide (Scheme II, Table I). Alkaline hydrolysis of ynenol lactones **1** results in the formation of allene acids **4**.⁸

(5) The iodo enol lactone **2** is treated with a terminal alkyne in the presence of a catalytic amount of cuprous iodide (0.05–0.1 equiv) and bis(triphenylphosphine)palladium(II) chloride (0.1–0.2 equiv) in triethylamine at 35 °C for 4–6 h. Yields vary from 44% to 66%. Deprotection of the trimethylsilylated ynenol lactones is achieved with AgNO₃/KCN in aqueous ethanol.

(6) In this study, only racemic ynenol lactones were tested. **1a**: ¹H NMR (CDCl₃) δ 2.8 (m, 2 H, H₃), 3.1 (d, 1 H, C≡CH), 3.1 (m, 2 H, H₄), 5.4 (ddd, 1 H, C=CH); IR (CHCl₃) 3300, 1810, 1660, 2100 cm⁻¹; MS, 122 (M⁺), 94 (M⁺ - CO). **1b**: ¹H NMR (CDCl₃) δ 0.9 (br t, 3 H, CH₃), 1.4 (m, 6 H, (CH₂)₃), 2.8 (m, 3 H, H₃, H₄), 3.1 (d, 1 H, C≡CH), 5.25 (m, 1 H, C=CH); IR (CHCl₃) 3300, 1805, 1655 cm⁻¹; bp 108–110 °C (1 mmHg); MS, 178 (M⁺), 111, 66, 28. **1c**: ¹H NMR (CDCl₃) δ 2.9 (d, 1 H, C≡CH), 2.5–3.2 (m, 5 H, PhCH₂, H₃, H₄), 5.2 (m, 1 H, C=CH), 7.2 (m, 5 H, Ar H); IR (CHCl₃) 3300, 1800, 1600, 2100 cm⁻¹; mp 64–65 °C; MS, 212 (M⁺), 145, 66, 91 (C₇H₇⁺).

(7) Coupling reactions carried out under these conditions proceed stereospecifically with retention of configuration. See: (a) Ratovelomanana, V.; Linstrumelle, G. *Tetrahedron Lett.* **1981**, 315–318. (b) Ando, T.; Vu, M.; Yoshida, S.; Takahashi, M. *Agric. Biol. Chem.* **1982**, *46*, 717–722.

(8) For example, **1a** (UV in H₂O: λ_{max} 229 nm (14 100 M⁻¹ cm⁻¹) at pH 10, 25 °C, gives **4a** (λ_{max} 220 nm (9600 M⁻¹ cm⁻¹) at a rate of 0.015 s⁻¹. In ¹H NMR, **1a** plus 1.8 equiv of KOH gives **4a** in situ (six-line ABX, δ_X 5.80, δ_A, δ_B centered at 5.37, J_{AX} + J_{BX} = 13.4 Hz). (a) Covey, D.; Robinson, C. H. *J. Am. Chem. Soc.* **1976**, *98*, 5038–5040. (b) Carlson, R.; Henton, D. J. *Chem. Soc., Chem. Commun.* **1969**, 674–675. (c) Abraham, R. J. "The Analysis of High Resolution NMR Spectra"; Elsevier: Amsterdam, 1971; pp 75–77.

Ultraviolet spectra during the course of hydrolysis of the terminal alkynes (**1**, R₄ = H) are isobestic (for **1a**, λ = 208 nm). In contrast, the terminally substituted alkynes (**1**, R₄ = alkyl) show biphasic, nonisobestic spectra during alkaline hydrolysis, probably due to the intermediate formation of the propargyl ketone **3** (R₄ = alkyl), which is only slowly isomerized to the allenone **4** (R₄ = alkyl).

We have focussed on the inhibition of human leukocyte elastase⁹ (HLE, E.C. 3.4.21.11.) because of the probable role of this enzyme in human disease.¹⁰ Ynenol lactone **1a** is an inhibitor of HLE (K_i = 5.3 μM, based on initial rate assays) that does not show measurable time-dependent inactivation, suggesting that acylation by **1a** (Scheme I, a → b), but not trapping (d → e), occurs.

Substitution at C-3 has a profound effect on potency. Compound **1c** shows time-dependent, irreversible inactivation. The rate of inactivation is saturable (K_{inact} = 4.1 μM, k_{inact} = 0.09 s⁻¹, giving an apparent rate at low concentrations of **1c** of 22 000 M⁻¹ s⁻¹). Inactivation is efficient; approximately two-thirds of all catalytic events lead to inactivation. Protection against inactivation by elastatin, an active-site directed, reversible inhibitor,¹¹ shows that **1c** is active-site directed. That the *tethered* allenone (Scheme I, d) is required for inactivation is demonstrated by the inability of exogenous allenone **4c** (100 μM; prepared in situ by alkaline hydrolysis of **1c**) to inactivate the enzyme. Inactivation by **1c** is irreversible, since inactivated enzyme shows no recovery of activity after gel exclusion chromatography (Sephadex G-50), with or without 2-mercaptoethanol present. Ynenol lactones **1b** and **1g** inhibit HLE with potencies comparable to **1c** (apparent second-order inactivation rates of 23 000 and 28 000 M⁻¹ s⁻¹, respectively).

On the other hand, the geminally substituted **1f** inhibits HLE 350-fold slower than **1c**. Terminal substitution of the alkyne **1d** significantly slows inhibition (1000-fold slower than **1c**). It is also noteworthy that the specificity constants (apparent k_{inact}/K_{inact}) for inhibition of the serine proteinases HLE, porcine pancreatic elastase, and bovine trypsin are, respectively, 22 000:730:17 for **1c** and 23 000:260:43, for **1b**.

Acyl enzyme formation by ynenol lactones has been demonstrated more directly with bovine α-chymotrypsin. Addition of a *catalytic* amount of chymotrypsin to **1a** at neutral pH results in ultraviolet spectra identical with those seen during alkaline hydrolysis of **1a**, showing that **1a** is a substrate and that **4a** is its accumulating product. With *stoichiometric* **1c** and chymotrypsin, proflavin (0.02 equiv) is rapidly displaced from the active site and rebinds in a time-dependent manner. The rate of this rebinding is a measure of product deacylation,¹² and these rates show a pH profile similar to other acylchymotrypsins.¹³ These and other results will be elaborated in future publications.

Acknowledgment. We thank Valerie Robinson for her expert

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NMR assistance and helpful discussions and Lynn Jacob for her many contributions.

Supplementary Material Available: Spectral characterization of 1f-1g and a description of HLE assay (1 page). Ordering of information is given on any current masthead page.

Electrochemistry in Near-Critical and Supercritical Fluids. 1. Ammonia

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We report electrochemical studies in near-critical and supercritical ammonia and the behavior of solvated electrons and *m*-chloronitrobenzene in this medium. Thermodynamic and solubility studies of supercritical solutions have been an active area of research.¹ We are interested in applying electrochemical techniques to near-critical and supercritical fluids to obtain a better understanding of the thermodynamics and kinetics of reactions, particularly homogeneous and heterogeneous electron-transfer reactions, that occur in these media and perhaps to utilize these solutions for electrosynthetic purposes. The high temperatures and pressures necessarily associated with electrochemically useful supercritical solutions necessitate the use of highly specialized cells and electrodes. In addition, the increased reactivity of species and the highly corrosive environment of supercritical ammonia limits the types of useful electrolytes and redox couples that easily can be studied as test systems.

As the critical temperature of a liquid is approached, the gaseous and liquid phases merge into a single, space-filling phase called a supercritical fluid. For ammonia, the critical point occurs at 133 °C and 112.5 atm; addition of low concentrations of electrolyte do not change these values appreciably. The characteristics that typify these fluids include decreased viscosities, densities, and dielectric constants, unusual changes in inter- and intramolecular forces, and altered solvation characteristics. A primary question of interest is whether electrochemical studies can be carried out in a supercritical fluid containing an electrolyte. We show here that electrochemical techniques, such as cyclic voltammetry and chronocoulometry, with near-critical and supercritical ammonia can be used to probe changes in redox potentials, electrogenerated product stability, and diffusion coefficients. Of the limited number of electrochemical studies that have been carried out on supercritical solutions, most have dealt with the corrosion of metals in contact with water.² To our knowledge, the only previous attempt at obtaining quantitative electrochemical information from supercritical NH₃ involved a two-electrode, constant-current, electrodeposition of silver.³

There are two primary experimental difficulties associated with these types of experiments: containment of a high-pressure, space-filling, corrosive fluid and design of electrode feed throughs that can withstand the supercritical environment and remain insulated from the walls of the cell. The base of the electrochemical cell we have fabricated is of 316 stainless steel; the

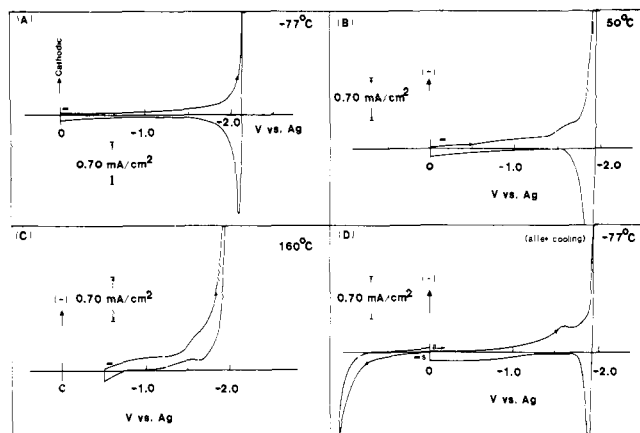


Figure 1. Cyclic voltammograms for generation and oxidation of solvated electrons in NH₃. (A) -77 °C, 0.2 M KI; (B) 50 °C, 20 atm, 0.2 M KI; (C) 160 °C, 252 atm, 0.1 M KI (supercritical); (D) -77 °C (after cooling from 160 °C), 0.2 M KI. Scan rate, 200 mV s⁻¹.

interior is cylindrical and has an internal volume of 75 mL. A detachable lid of the same material is bolted to the base by means of a flange arrangement incorporating a diamond-shaped copper gasket. Three electrodes, supported by standard Swagelok fittings, pass through the lid. Provisions are also made for evacuation, filling, sample addition, and a rupture disk. All electrode feed throughs consist of tungsten wire passed through a commercially available graded glass to Kovar seal. Construction of the feed through is completed by sealing the glass to the wire. The working electrode is a disk-shaped cross section of a tungsten wire, and the counter and quasi-reference (QRE) electrodes consist respectively of platinum and silver wires soldered onto the tungsten. These electrodes have withstood pressures of 340 atm at 160 °C.

A typical experiment involved evacuating the cell, to which electrolyte had previously been added, followed by the addition of sufficient dry ammonia under vacuum line conditions to generate the desired pressure and density at a specified temperature. The cell was isolated from the vacuum line by means of a high-pressure valve, and removed to an armored autoclave for heating above the critical temperature (~140 °C) of the electrolytic solution. To probe changes in the available potential range of NH₃ and the stability of solvated electrons (e_s⁻) as a function of temperature, cyclic voltammetric scans were employed (Figure 1).⁴

As the temperature increased, the polarizable range of the ammonia solution decreases as evidenced by a positive shift of the solvated electron peak and a negative shift of the anodic background (vs. QRE). A small shoulder preceded the onset of solvated electron production at higher temperatures which persisted when the solution was cooled back to -77 °C (curve D). A similar wave was observed at -77 °C in single-compartment glass cells and thus is probably associated with the reduction of a species generated at the counter electrode. Above the critical temperature (curve C) generation of solvated electrons is still observed, although on the voltammetric time scale little, if any, oxidation is seen upon scan reversal. This can be attributed to a decrease in the stability of the solvated electrons, probably by reaction with NH₃ at the higher temperatures.⁵ Upon cooling the solution to -77 °C, the system returns to essentially its initial condition, demonstrating no extensive contamination or irreversible changes in the solution. The apparent shift in potential of the solvated electron peak

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